

Sustainable Food Technology

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Sustainability Spotlight Statement

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Being a sustainable alternative to traditional animal-based foods, plant-based foods are increasingly used in human diets. However, inferior techno-functional properties of plant-based ingredients, which are causing inferior sensory attributes in foods, are hindering the expected growth of plant-based foods in the commercial market. The high-oil-containing powder presented in this manuscript has better functional properties, such as dispersibility and reconstitutability, making it suitable for enhancing sensory attributes of various plant-based products such as plant-based milk, cream, cream cheese, cheese, etc. The oil core present in oil globules can also be used as a vehicle for delivering hydrophobic health-promoting compounds, helping enhance the well-being of humans. We understand that this product/process would help achieve SDGs 3, 9 and 13.



1 **Innovative formulations using spray-drying technology for plant-based high-oil**
2 **powders: physicochemical and micro-structural analyses**

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10

11 **ABSTRACT**

12 Plant-based ingredients, which are considered sustainable sources, are increasingly
13 used to produce food alternatives to animal-origin products. However, despite being
14 considered a sustainable option, the wider acceptance of plant-based alternative foods
15 is poor. The major reasons are the lack of desirable functionalities in plant-based food
16 ingredients and the inferior sensory attributes of prepared foods and the lack of desirable
17 functionalities in plant-based food ingredients compared to their animal-based
18 counterparts. To fulfil this gap, this study focuses on the production and characterization
19 of plant-based high-fat powder with enhanced functionalities, which could serve as an
20 alternative ingredient to the dairy-based cream powder in the food manufacturing sector.
21 Plant-based high-oil powders containing 20% and 40% total oil were prepared from corn
22 oil emulsion having a mean oil globule size of 0.47 μ m and 0.75 μ m by spray drying.
23 Formulations used a water-soluble fraction of mung bean protein isolate as an emulsifier



24 and maltodextrin as a wall material. The physicochemical analyses of the powders
25 revealed that the powder prepared from corn oil emulsion with a mean fat globule size
26 (D[4,3]) of 0.47 μ m and 20% oil content had a lower angle of repose, higher bulk density
27 and lower free oil content than other high-oil powder samples. The confocal laser
28 scanning microscopy (CLSM) and scanning electron microscopy (SEM) images also
29 showed that powders prepared from smaller fat globules were individual, with low
30 surface oil coverage compared to the powders prepared from larger fat globules. This
31 study highlighted the suitability of plant-based sources for developing high-oil powders
32 that could find potential applications in creating valuable food products.

33

34 **KEYWORDS**

35 Plant-based high-oil powder, emulsion, mung bean, spray drying, microstructure

36 **1. Introduction**

37 High-fat/oil powders are widely used in food product development because of their
38 nutritional, textural, and flavor attributes. These powders, containing a high percentage of
39 fats, from 40%, find applications in bakery products, soups, sauces, processed meats, creamy
40 condiments, evaporated milk, infant formulas, cheese, coffee, and tea. Owing to their low
41 water activity and moisture content, these powders have a long shelf life and are convenient
42 for transportation, storage, and use. These powders are manufactured at the commercial scale
43 using spray drying to ensure they possess essential instant properties such as wettability,
44 sinkability, solubility and dispersibility. While high-fat powders have a long shelf life due
45 to their low water activity and moisture content, they are more prone to fat oxidation and
46 particle caking during storage. The fat oxidation can be prevented by encapsulating it in a
47 wall material before spray drying [1-3].



48 The first step for the encapsulation is to prepare the stable emulsion, which can be done
49 through homogenization techniques such as high-pressure homogenization, ultrasonication,
50 and microfluidization [4]. Among these, ultrasonication forms a nano emulsion with the least
51 energy and offers better stability [5].

52 Different synthetic (polysorbates, acetylated mono and diglycerides, carboxymethyl
53 cellulose etc.) and natural (caseinates, whey proteins, lecithin, plant proteins, etc.)
54 compounds can be used as food emulsifiers [6]. Among these, proteins are the best
55 contenders for use in emulsion preparation due to their amphiphilic nature. In the present
56 context, the most widely used protein-based emulsifiers are caseinates and whey proteins.
57 Several studies have been conducted on oil-in-water emulsions prepared using caseinates,
58 whey proteins or their mixture in the last few decades [7-10]. These emulsions have been
59 successfully used to prepare dairy-based high-fat powder, such as cream powder. However,
60 the research on applying plant proteins as an emulsifier for preparing plant-based high-fat
61 powder is limited [11, 12]. While soy protein is abundant and possesses excellent emulsifying
62 properties, the regulatory concerns over allergen status have necessitated researchers to find
63 novel alternatives [13]. Mung bean protein, particularly its water-soluble fraction, has
64 recently gained attention for its various techno-functional properties, including foaming,
65 solubility, emulsifying capacity, and water and oil absorption capacity [14]. In the studies
66 conducted by Brishti, Zarei et al. [15] and Du et al. [16], the emulsifying, water absorption
67 and gelling properties of mung bean protein isolates were more desirable compared to other
68 plant proteins. Furthermore, the bland taste and mild odour of mung bean protein make it
69 more preferable when compared with soy protein [17].

70 The second step in encapsulation involves selecting the appropriate wall material and
71 understanding its properties [18]. The wall material can be proteins and water-soluble
72 carbohydrates such as maltodextrin and gums. The wall material used in high-fat powder



73 embeds the fat globules in its continuous network. However, if the fat content in the emulsion
74 is increased without increasing the wall materials proportionally, a thinner protective layer
75 is formed around the fat globules. This results in an increase in free fat content in the
76 encapsulated material (i.e. oil phase) [10]. The free fat content is crucial for ensuring the
77 functionalities of food powders, such as high-fat powder. In the studies conducted by Drusch
78 and Berg [19] and Sarkar et al.[20], there was an increase in the surface fat content and
79 number of fat globules close to the surface of the powder particles with an increase in total
80 fat content in the powder. The free fat content in a spray-dried powder represents the surface
81 fat, capillary fat, and outer layer fat that are easily extractable in the solvent [19]. The amount
82 of free fat content influences the functional properties of the powder, such as oxidative
83 stability, flowability, wettability, and reconstitutability [21]. Higher free fat content leads to
84 lower shelf life because of oxidative rancidity [8].

85 Mohammed et al.[22] reported that high water-soluble and low viscous wall materials are
86 ideal for the preparation of spray-dried powder [22]. Maltodextrin is such an example, which
87 is an inexpensive, flavourless starch hydrolysate that can protect the encapsulated oil phase
88 from oxidative degradation. Furthermore, it is highly soluble in cold water, which aids in the
89 process of emulsification[22]. In a study conducted by Munin and Edwards-Lévy [23], high
90 encapsulation efficiency of oil was achieved when a blend of maltodextrin and sodium
91 caseinate was used as wall material. The ability of the wall material to keep the core material
92 intact determines the encapsulation efficiency of the material. In general, the proportion of
93 fat that cannot be extracted as free fat gives the encapsulation efficiency of the wall material
94 in fat powders [24].

95 Powder properties are also affected by the melting point of the fat in the core. Fat powder
96 containing encapsulated fat with a low or high melting point tends to have lower free fat
97 content compared to fat with an intermediate melting point. The reason behind this is the



98 presence of partially crystalline fat in the liquid oil phase. The partial crystalline phase tends
99 to break the protective film that surrounds the partially liquid droplets, leading to fat leakage
100 and poor encapsulation [10, 25, 26]. Furthermore, the encapsulation efficiency is also
101 affected by the droplet size of the emulsion. Smaller droplet size in the emulsion generally
102 leads to less free fat content in the powder or greater encapsulation efficiency. Upon
103 decreasing the mean droplet size from 1.2 μm to 0.5 μm , a seven-fold decrease in surface fat
104 content in the fat powder was observed by Danviriyakul et al.[27]. Similar results were
105 reported by Jafari et al.[9] in the study of fish oil powders. The author reported a decrease in
106 free oil content from 1.27% to 0.69% when the average fish oil droplet size in the emulsion
107 was reduced to 0.28 μm from 5.9 μm [9].

108 While the market for plant-based food products is continuously increasing, there is a need
109 for the development of plant-based high-fat powder that can replace dairy cream powder.
110 Commercial dairy cream powder is known for better functionalities such as reconstitutibility
111 and dispersibility. Numerous studies have been carried out on powdered emulsions; however,
112 the materials (i.e. oil phase, emulsifiers, wall material) are either entirely animal-based or a
113 combination of animal and plant-based, or plant-based with limited solubility or lower fat
114 content [28]-[29]. To the best of our knowledge, very limited studies have been done to date
115 that produced plant-based spray-dried high-fat (>40% w/w) yet highly soluble powder. The
116 product in this study is aimed to be an equivalent alternative to dairy cream powder.
117 Therefore, this study investigated the various physicochemical properties of the high-fat
118 powder prepared using only plant-based materials (i.e. oil phase, wall material and
119 emulsifier).

120 **2. Materials and methods**

121 *2.1. Materials*



122 Corn oil (Reinna Brand) was purchased from a local supermarket in Perth, WA,
 123 Australia. Mung bean protein isolates were purchased from Bulk Powders (Australia),
 124 and maltodextrin (unflavored) was purchased from nutricost.com, USA. Analytical
 125 grade petroleum ether (BP= 40-60 °C) was purchased from Sigma-Aldrich, Bayswater,
 126 Victoria, Australia.

127 2.2. Methods

128 2.2.1 Experimental design

129 Table 1 shows the formulation used for preparing oil powder containing 40% and
 130 20% oil, and Table 2 shows the oil percent and the oil globule size for the four different
 131 samples prepared in this study.

132 **Table 1** Formulations used for the preparation of oil powder containing 40% and 20% oil.

Ingredients	Weight (g)	Weight (g)
Corn Oil	44	22
Supernatant (containing 0.3% protein)	250	250
Maltodextrin	67	89
Addition of water to dissolve wall material (maltodextrin)	150	150
Final Fat percentage (%) in spray-dried powder	40	20
Total	511	511
Solids in total mix, including oil phase (%)	23	23

133

134

135

136

137



138 **Table 2**

139 Description of the spray-dried powder samples

Total oil content (%) in spray-dried powder	Size of the fat globule in emulsion before spray drying (μm)	Sample Code
20	0.47	PS0.47-20
40	0.75	PS0.75-40
20	0.75	PS0.75-20
40	0.47	PS0.47-20

140 Note: The average size of the oil droplets of the emulsion collected after atomization
141 remained the same as before atomization.

142
143 *2.2.2. Sample preparation.*

144 *2.2.2.1. Mung bean protein isolate as emulsifier*

145 The emulsifier used in the study was prepared by modifying the method used by Wei
146 et al.[30]. Four g of mung bean protein isolate (protein content, 85.35g/100g powder)
147 was soaked in 296 g of deionized water. The soaked mung bean protein isolate was kept
148 overnight in a refrigerator. It was then centrifuged at 4700 rpm (Eppendorf 5810 R,
149 Germany) for 30 min, and the supernatant was carefully transferred into a beaker. It was
150 analyzed for protein content using the standard Kjeldahl method using a 6.25
151 multiplication factor. Further details of protein content determination are not elaborated
152 in this manuscript. The protein present in the supernatant was used to prepare the
153 emulsion. The aim of this work was to study the effect of emulsion droplet size on the
154 physicochemical properties of the high-fat powder. Only supernatant (completely
155 soluble fraction) was used to nullify the effect of undissolved protein particles on the
156 average size of the emulsion droplet while measuring via particle size analyzer. In our
157 preliminary trials, we observed that it was impossible to completely dissolve the mung-



158 bean protein powder particles, which were reflected in the particle size results.

159 2.2.2.2. *Emulsion preparation*

160 For emulsion preparation, the formulation in Table 1 was used. To prepare an oil-in-
161 water emulsion (E1) for spray-dried powder containing 40 % (w/w) oil, 44 g of oil was
162 mixed with 250 g of supernatant (0.3 %w/w protein content). The mixture was pre-
163 emulsified for 5 min at 17500 rpm using an Ultra-turrax homogenizer (IKA T18 basic,
164 Germany). Then, it was immediately homogenized using an ultrasonicator (VCX750,
165 Sonics & Materials Inc., Newtown, USA) for 8 min and 16 min at 27°C to obtain two
166 different fat droplet sizes (i.e. D[4,3] of 0.75 µm and 0.47 µm, respectively). Similarly, to
167 prepare emulsion (E2) for spray-dried powder containing 20% (w/w) oil, 22 g of oil was
168 mixed with 250 g of supernatant. Then Ultra-turrax homogenizer was used for 5 min at 17500
169 rpm and then ultrasonicated for 4 min and 12 min at 27°C to obtain particle sizes D [4,3] of
170 0.47 µm and 0.75 µm, respectively. Ultrasonication was carried out using the frequency of
171 20kHz, 600 W power, and 50% amplitude in continuous mode (without pulse) to prepare E1
172 and E2. The pre-emulsified sample was kept in an ice bath during the time of ultrasonication
173 to prevent an excessive rise in temperature. The prepared emulsions were analysed for oil
174 globule size and stored in refrigerated conditions.

175 2.2.2.3. *Wall material*

176 The wall material solutions were prepared by dissolving 67 g of maltodextrin in 150 g of
177 deionized water for spray-dried powder containing 40% (w/w) oil and by dissolving 89 g of
178 maltodextrin in 150 g of deionized water for spray-dried powder containing 20% (w/w) oil
179 (Table 1).

180 2.2.2.4. *Spray drying of the emulsion*

181 The emulsion and wall material were mixed using an overhead stirrer at 150 rpm for 3
182 min. Then, it was spray dried using a spray dryer (Buchi Mini Spray Dryer B-290, Buchi Co.



183 Switzerland). Feeding of the emulsion was done at 45 °C with a feeding rate of 5 mL/min
184 through a 0.7 mm diameter two-fluid nozzle. The compressed air pressure going to the
185 atomiser was set at 0.6 MPa. The inlet and outlet temperatures of the drying air were
186 maintained at 170 °C and 70 °C, respectively. The powdered samples were collected from
187 the cyclone separator and the collecting vessel by using a soft brush into airtight 70 mL sterile
188 containers. The powder from the drying chamber was not used for analysis. The sample
189 containers were kept in resealable bags and stored at refrigeration temperature until further
190 analysis. Each batch of freshly prepared powdered samples was analysed within a week.

191 2.3. Product characterization

192 2.3.1. Size measurement

193 The measurement of average oil globule size in the emulsion before spray drying
194 was carried out using a particle size analyzer (Malvern Mastersizer 2000, Malvern
195 Instruments Ltd. Worcestershire, UK). Deionized water was used as a dispersant, and
196 the emulsion was added dropwise to the deionized water using a dropper until a laser
197 obscuration of 10-10.5% was obtained. The refractive index values used for deionized
198 water (dispersant) and oil (dispersing material) were 1.33 and 1.47, respectively. The
199 absorption index was set at 0.01.

200 2.3.2 Moisture content and water activity

201 The moisture content of the oil powder was measured using the hot air oven method.
202 The oil powder was heated at 105 °C in a hot air oven until a constant weight was
203 reached. Water activity was measured using a water activity meter (AQUALAB 4,
204 METER Group Inc., USA) using approximately 3 g of sample to fill the base of the
205 sample holding cup.

206 2.3.3 Free oil content

207 Free oil content of the oil powder was determined by modifying the process



208 described by Schuck et al.[31]. Briefly, 5 ± 0.5 g of the oil powder was weighed in a
209 conical flask, and 100 mL of petroleum spirit was added to the flask. The conical flask
210 was then kept in an electric shaker for 15 min with gentle shaking, enough to ensure
211 adequate mixing without creating high turbulence. The solution was then filtered into a
212 pre-weighed dry round-bottom flask using Whatman filter paper 41. Then, 50 mL of
213 petroleum spirit was poured into the residue in the conical flask and was filtered into the
214 round-bottom flask. The solvent was evaporated off using the rotary vacuum evaporator
215 at 60°C. Then, the round-bottom flask was dried in the hot air oven at 105°C for 1 h.
216 The flask was then kept in a desiccator to cool down, and the dried weight was measured
217 as extractable fat. The following equation was used to calculate the percentage of free
218 oil content:

$$219 \text{ Free oil content (\%)} = (\text{Extractable oil (g)}/\text{Total oil in powder (g)}) \times 100.$$

220 The residue was left to dry in the fume hood overnight and collected into an airtight
221 container for the analysis of solvent-washed high-oil powder.

222 2.4.4. *Oil globule size of reconstituted powder*

223 The reconstitution of the spray-dried fat powder and the solvent-washed fat powder
224 was carried out as described by Hogan et al.[32]. For the reconstitution, 0.5 g of the
225 powder was dissolved in 150 mL of deionized water. The solution was gently stirred
226 using an overhead stirrer for 30 min at room temperature (25-28 °C). Then the oil globule
227 size was measured using Mastersizer as described in section 2.3.1. The oil globule size
228 of the reconstituted spray-dried powder and solvent-washed reconstituted powder was
229 compared with the mean oil globule size of the parent emulsion. Parent emulsion
230 indicates the emulsion before spray drying.

231 2.4.5. *Angle of repose*

232 The angle of repose of the spray-dried powder and the solvent-washed powder was



233 measured as described by Kim et al.[33]. The powder was poured through a funnel into
234 a petri plate (diameter: 80 mm, radius: 40mm). The maximum height (mm) of the
235 powder was recorded after it fully covered the base. It was then calculated using the
236 following equation:

$$\theta = \tan^{-1} \frac{\text{radius of Petri plate}}{\text{height}}$$

238 2.4.6. Bulk density

239 Briefly, the fat powder was filled into a 10 mL measuring cylinder. The weight of
240 the powder was then divided by the volume of the cylinder to calculate the bulk density
241 [18].

242 2.4.7. Confocal laser scanning microscopy (CLSM)

243 The microstructure of the oil powder was visualized using a confocal laser scanning
244 microscope (Nikon A1+, US). The oil powders were dyed with Rhodamine B for protein
245 and Nile red for fat, 10 min before imaging. The dyes were prepared by dissolving 5 mg
246 of each dye powder in 50 mL of polyethylene glycol 400. To stain the powder, 20 mg of
247 each of the dyes was added to 0.1 g of powder and gently mixed. A thin layer of stained
248 powder was placed on a slide and gently pressed with a coverslip. The powders were
249 then observed with a 63X water-immersion objective. For the excitation of Nile Red and
250 Rhodamine B, lasers with wavelengths 488 nm and 555 nm were used, respectively [10].

251 2.4.8. Scanning electron microscopy (SEM)

252 The surface morphology of the oil powders was studied using SEM (VEGA3
253 TESCAN, Brno, Czech Republic). To visualize the powder morphology, the samples
254 were mounted on the SEM stubs, and a sputter was used to coat them with gold. To
255 ensure the samples are placed securely in the stubs, double-sided adhesive tape was used
256 as described by Shivakumar et al.[18]. The samples were then examined with 5000x



257 magnification.

258 2.5 Statistical Analysis

259 Two-way analysis of variance with no blocking was carried out using GenStat (12th
260 Edition, VSN International Ltd.); the significant difference was considered at a 5% level
261 of significance. To determine whether the sample means were significantly different or
262 not, LSD and interaction effects were obtained. All the analysis was done in triplicate.

263 3. Results and discussion

264 3.1 Water activity and moisture content

265 The mean water activity of the spray-dried oil powders was measured to be
266 0.13 ± 0.01 , 0.14 ± 0.01 , 0.15 ± 0.01 and 0.16 ± 0.01 for PS0.75-40, PS0.4-20, PS0.75-20
267 and PS0.47-40, respectively. There was no significant difference ($p>0.05$) in the values
268 of water activity among the powders. Likewise, the mean moisture content of the
269 powders was 4.48 ± 0.24 , 4.46 ± 0.21 , 4.82 ± 0.22 and 4.52 ± 0.01 % for PS0.75-40, PS0.47-
270 40, PS0.75-20 and PS0.47-20, respectively. There was no significant difference ($p>0.05$)
271 between the mean moisture content of the powdered samples. A similar result was also
272 obtained by Dhungana et al. [10] in the spray-dried high-fat powder that was prepared
273 from a cream emulsion. The authors also reported no significant difference in the mean
274 moisture content and water activity of the powdered samples having a particle size
275 ranging from $0.21\ \mu\text{m}$ to $1.42\ \mu\text{m}$ with fat content ranging from 35% to 75 %. Similarly,
276 no difference in moisture content and water activity, even when the inlet temperature of
277 the spray dryer varied between 150, 170 and 190 °C was observed by Himmetagaoglu
278 and Erbay [34] in their study of fat powders. The drying conditions used for all the
279 powders (i.e. PS0.47-20, PS0.47-40, PS0.75-20 and PS0.75-40) were the same, which
280 could be the reason for no significant difference in the mean values of moisture content
281 and water activity.



282 3.2 Bulk density

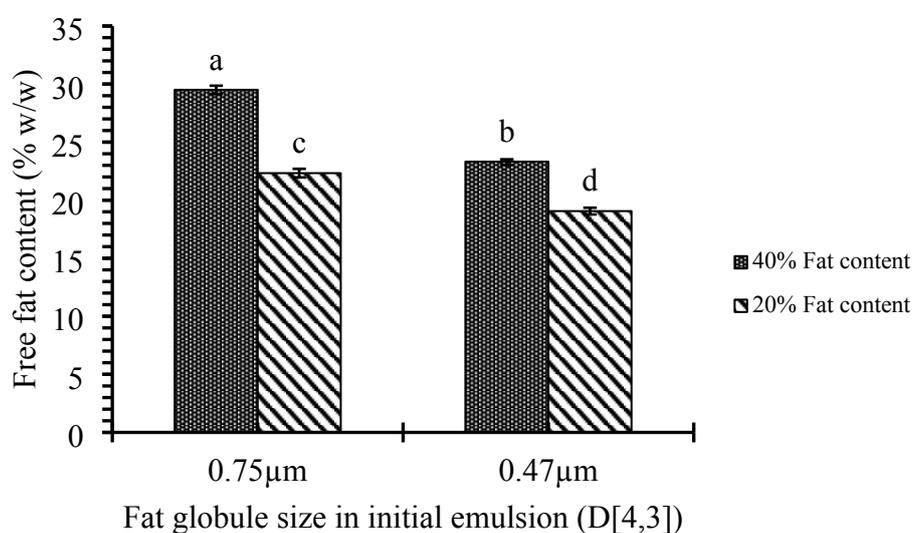
283 The average bulk density of the oil powders was 0.236 ± 0.004 , 0.239 ± 0.006 ,
284 0.246 ± 0.005 and 0.252 ± 0.005 g/cm³ for oil powders PS0.75-40, PS0.47-40, PS0.75-20
285 and PS0.47-20, respectively. A significant effect ($p<0.05$) of oil content on the bulk
286 density of the fat powder was observed when two-way ANOVA was carried out. There
287 was an increase in the bulk density of the fat powder with a decrease in total fat content
288 in the powder. A similar result was obtained by Zhang et al.[35], where the bulk density
289 of soya bean oil powder increased from 0.769 g/cm³ to 0.916 g/cm³ when the oil content in
290 the powder was decreased to 10% from 20%. Likewise, Domian et al.[36] also reported an
291 increase in bulk density (0.282 g/cm³ to 0.354 g/cm³) of spray-dried emulsion when the fat
292 content dropped (55% to 40%). This trend is primarily attributed to the higher density of
293 maltodextrin compared to oil. Besides, the powders with higher fat content had characteristic
294 dents, which resulted in lower bulk density [36]. The results obtained showed a direct
295 relationship between the morphology of powders and different wall thicknesses. High-oil
296 powders with low maltodextrin content led to thinner shell-wall formation around the oil
297 droplets, resulting in powder particles with irregular shapes. These irregularly shaped powder
298 particles required more space, leading to a lowered bulk density of the powder. On the other
299 hand, oil powders with thicker shell-walls have relatively small and uniform sizes, which
300 resulted in higher bulk density [35].

301 3.3 Free fat content

302 Based on total oil weight in powder, the powdered corn oil samples PS0.75-40,
303 PS0.47-40, PS0.75-20 and PS0.47-20 had a mean free oil content of 29.51 ± 0.35 ,
304 23.32 ± 0.28 , 22.33 ± 0.37 and 19.60 ± 0.20 %, respectively (Fig. 1). The results of ANOVA
305 indicated a significant effect ($p<0.05$) of total oil content, oil globule size and their
306 interaction term (oil globule size \times total oil content) on the mean free oil content of the



307 powder. There was a significant ($p < 0.05$) decrease in average free oil content with the
 308 decrease in mean oil globule size of the initial emulsion and a drop in total oil content
 309 in the final powder. Similar results were obtained by Dhungana et al.[10] in spray-dried
 310 cream emulsions. The authors reported an increase in free fat content from 1.8% to
 311 75.6% when the fat globule size of the parent cream emulsion was increased from 0.21
 312 μm to 1.41 μm , and the total fat content in the powder was increased from 35% to 70%,
 313 respectively.



314
 315 **Fig. 1.** Free fat content (g/100 g oil in powder or %w/w) of the spray-dried corn oil
 316 powder prepared from parent emulsion size with oil globule size of 0.75 μm and 0.47
 317 μm .

318 The increase in surface oil content from 45.3% to 48.9% was also reported by Hogan
 319 et al.[32] when the mean droplet size of the oil emulsion was increased from 0.41 μm to
 320 1.41 μm . The oil-to-sodium caseinate ratio in their experiment was 1. Similarly, in the
 321 study of spray-dried encapsulated fish oil powders in which whey protein concentrate
 322 was used as a stabilizer, Jafari et al.[9] observed an increase in free fat content from 690
 323 mg/100g to 1270 mg/100 g when the mean droplet size of the fish oil emulsion was



324 increased from 0.28 μm to 5.9 μm . Furthermore, an increase in free fat content with an
325 increase in droplet size in the emulsion was also reported by Danviriyakul et al. [27].
326 The free fat content of the spray-dried milk fat prepared from the emulsion with a mean
327 droplet size of 0.5 μm was 2%, whereas the milk fat powder prepared from the emulsion
328 with a mean droplet size of 1.2 μm was 13.2%. The reason behind this could be because
329 of the ability of the smaller fat globules to disperse more uniformly within the wall
330 matrix during the process of spray drying. An even dispersion of fat globules within the
331 wall matrix decreases the chances of being washed away during the process of solvent
332 washing. Furthermore, the migration of larger fat globules to the outer part of the sprayed
333 droplets is comparatively faster than that of smaller fat globules. Therefore, bigger fat
334 globules tend to remain on the surface of the powder particles, which increases their
335 chances of being easily washed away by the solvent [9]. Moreover, the larger fat
336 globules are highly sensitive to mechanical stress and can easily break down during the
337 process of spray drying. These ruptured fat globules remain on the surface of the powder
338 and can easily get washed away with the solvent, resulting in higher free fat content [10].

339 The increase in free fat content with the increase in total fat in the final powder was
340 also reported by Hogan et al.[32]. A drop in the surface oil content to 10.85% from
341 81.2% was reported by the authors when the ratio of oil to sodium caseinate was
342 decreased from 3 to 0.25. Since, with an increase in total fat content, there is a decrease
343 in the amount of wall material used (Table 1). This can lead to the formation of thinner
344 interfacial membranes around the fat globules. This results in a higher chance of the fat
345 globules being broken down and subsequently washed away during solvent extraction.
346 Furthermore, an increase in the amount of wall materials helps keep the fat globules far
347 away from each other, which ultimately decreases the chances of re-coalescence and
348 fusion of the fat globules. This can lead to lower free fat content since the fat globules

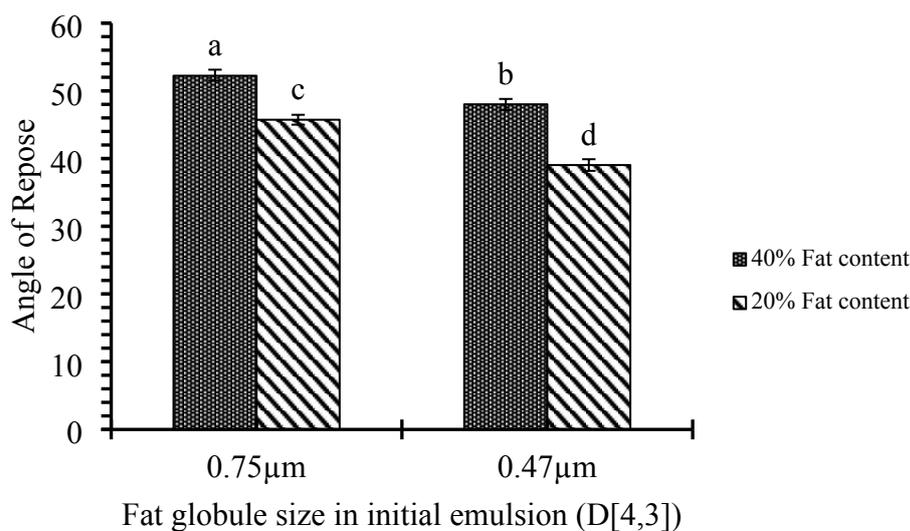


349 are embedded and dispersed evenly within the wall matrix [10, 32].

350 3.4 Angle of repose

351 The angle of repose of the high-oil powders, PS0.75-40, PS0.47-40, PS0.75-20 and
352 PS0.47-20 was $52.23\pm 0.89^\circ$, $48.01\pm 0.73^\circ$, $45.73\pm 0.79^\circ$ and $39.04\pm 0.86^\circ$ respectively
353 (Fig. 2). Two-way analysis of variance indicated a significant effect ($p < 0.05$) of total fat
354 in powder, mean oil globule size in parent emulsion and their interaction (total fat \times
355 content fat globule size) on the angle of repose of the powder. An increase in mean oil
356 globule size in the emulsion and an increase in total oil content in the powder resulted
357 in an increase in the angle of repose of the high-oil powder. Similar results were obtained
358 by Kim et al.[33] in dairy powders. The authors reported an increase in the angle of
359 repose of the dairy powders (skim milk, whole milk, and cream) with different fat
360 content. Furthermore, the authors also reported an increase in the angle of repose for
361 each sample when the fat globule size was varied for each level of fat content. The angle
362 of repose is an indication of the flow properties of the powders. A lower angle of repose
363 indicates higher flowability of the powder and vice versa. The increase in the angle of
364 repose could be due to an increase in the free oil content, which increased with an
365 increase in oil globule size and oil content in the powder, that acts as a connecting bridge
366 between the powder particles, leading to the clumping of the powder particles and
367 decreasing their flowability [33].





368

369 **Fig. 2.** Angle of repose of the spray-dried high-oil powders having two different fat
 370 contents (i.e. 40% and 20%) and two different oil droplet sizes (i.e. 0.75 μm and 0.47
 371 μm) in the parent emulsion.

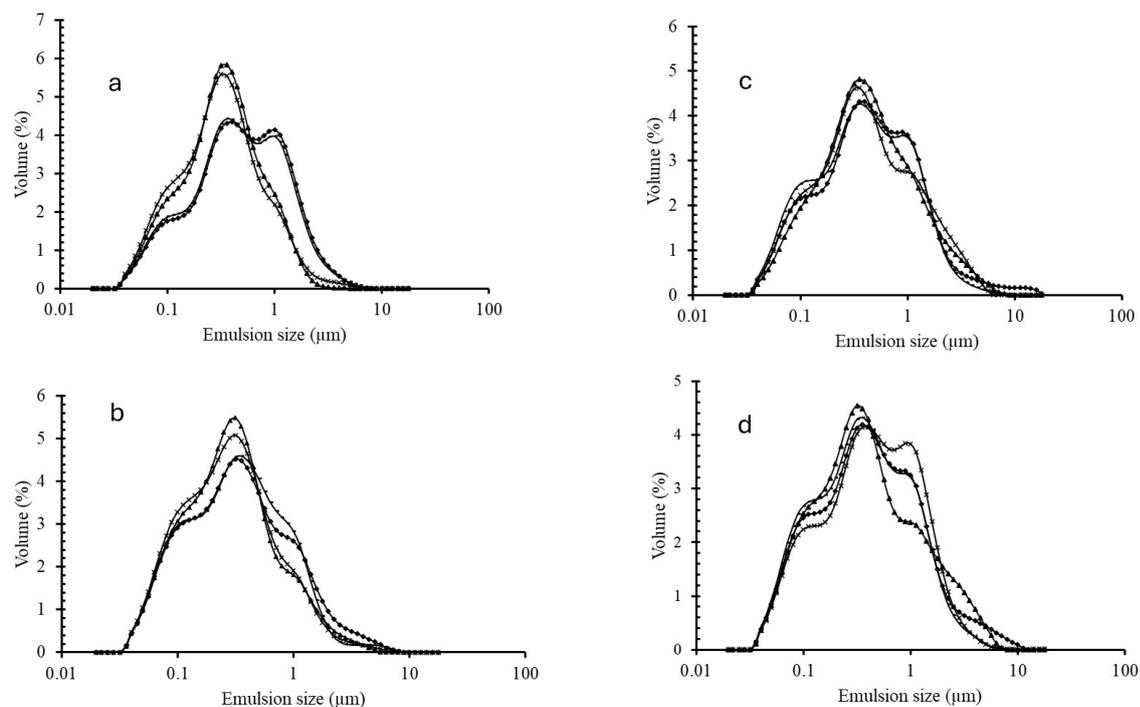
372 *3.5 Oil globule size of the reconstituted high-oil powder and solvent-washed*
 373 *reconstituted powder.*

374 The size distribution plots of the initial emulsion, after the addition of maltodextrin
 375 and after reconstitution of spray-dried powder and of the reconstituted emulsion prepared
 376 from solvent-washed powder are given in Fig.3. In all cases, the mean oil globule size of the
 377 corn oil emulsion remained almost the same after the addition of the maltodextrin.
 378 Maltodextrin was added after its solubilization. However, there was an increase in the mean
 379 oil globule size of the reconstituted powder samples. Similar results were reported by Ixtaina
 380 et al.[24]. The authors reported an increase in the mean fat globule size of the powder
 381 compared to the parent emulsion. Among the powders prepared from the same-sized parent
 382 emulsions, the extent of oil globule size increment in their reconstituted solutions was higher
 383 for the powder with higher oil content. This could be due to higher free oil on the powder
 384 with higher oil content, which was noticeable during size measurement as bigger oil globules



385 (coalesced) upon reconstitution.

386 In the case of reconstituted emulsion prepared from solvent-washed powder, the
387 mean droplet size for the oil powders prepared from the parent emulsion with a mean size of
388 $0.47\ \mu\text{m}$, there was only a slight drop in the mean oil globule size (Fig. 3a & 3b). However,
389 the mean oil globule size was lower for the reconstituted emulsion prepared from the parent
390 emulsion with a mean size of $0.75\ \mu\text{m}$ (Fig. 3c & 3d). A similar drop in the average fat
391 globule size compared to reconstituted emulsion was also reported by Dhungana et al.[10].
392 This could be due to the presence of less stable free oil droplets on the surface of the powder
393 prepared from larger fat globules in the parent emulsion.



394
395 **Figure 3.** a) Oil globule size distribution plot ($0.47\ \mu\text{m}$ and 40% oil); b) Oil globule size
396 distribution plot ($0.47\ \mu\text{m}$ and 20% oil); c) Oil globule size distribution plot ($0.75\ \mu\text{m}$ and
397 40% oil); d) Oil globule size distribution plot ($0.75\ \mu\text{m}$ and 20% oil). Initial emulsion ($\text{---}\blacktriangle\text{---}$),
398 emulsion after addition of maltodextrin ($\text{---}\times\text{---}$), reconstituted emulsion from high-oil powder
399 ($\text{---}\bullet\text{---}$), reconstituted emulsion from solvent-washed powder (---)



400

401 The plots in Fig. 3 show the change of oil globule size distribution from unimodal in the
402 initial emulsion to multimodal or bimodal in the reconstituted powder, which could be due
403 to coalescence that might have occurred during reconstitution [24, 32]. However, the oil
404 globule size distribution of reconstituted powder is not significantly skewed to the larger
405 globule size zone, as is visible in the plots. This indicates that the high-fat powder prepared
406 for this study has very good reconstitutability. Ideally, if the powder surface is free of surface
407 oil or if there is no breakage of oil globules during spray drying and storage, the reconstituted
408 emulsion should have a similar oil globule size and distribution as the parent emulsion. This
409 is the main achievement of this study, to produce a high-oil plant-based powder with better
410 reconstitutability. The reconstitutability is one of the crucial powder properties that governs
411 its functionality in product development. Similar fat globule size distribution plots were also
412 obtained by Dhungana et al.[10] in the study of spray-dried high-fat powder prepared from
413 the dairy cream emulsion.

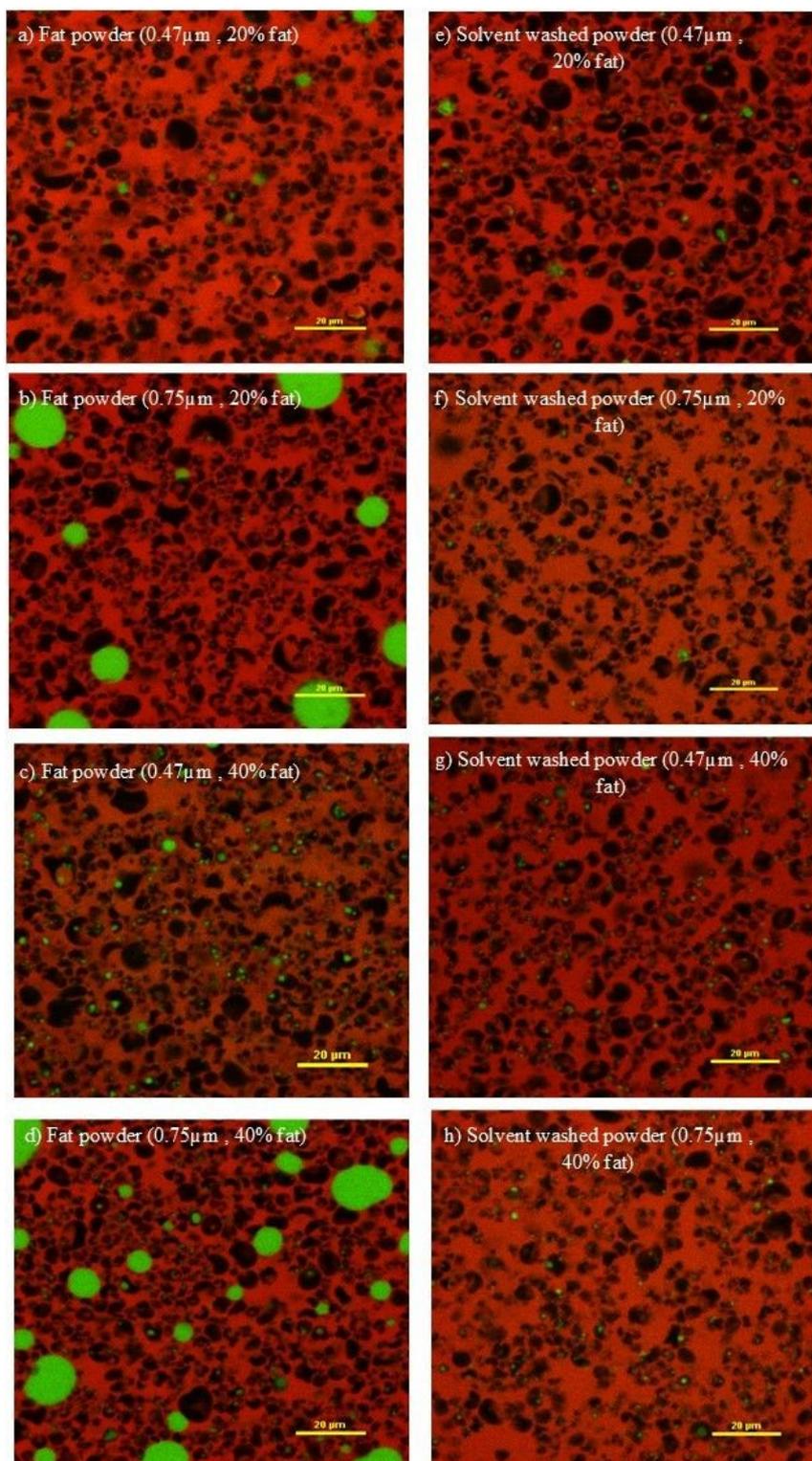
414 3.6. Confocal laser scanning microscopy (CLSM) of the corn oil powders

415 The CLSM images of high-oil powders and solvent-washed high-oil powders are
416 given in Fig. 4. The green color in these images denotes oil. The images show a higher
417 proportion of oil on the surface of the powder with the increase in the average fat globule
418 size of the corn oil emulsion. Furthermore, on lowering the total oil content in the powder
419 prepared from corn oil emulsion having the same mean oil globule size (0.47 μm and
420 0.75 μm), a decrease in the proportion of oil present on the surface of the oil powder was
421 observed. The images obtained are comparable to those obtained by Dhungana et al.[10]
422 in the study of cream powders and Kosasih et al.[37] in the study of spray-dried whole
423 milk powders. The observations made through images are also supported by the values
424 obtained for free oil content (i.e. the oil powder with the highest amount of free fat



425 content has the highest proportion of surface oil in the image and vice versa) as presented
426 in Fig. 1. The CLSM images (Fig 4e, 4f, 4g & 4h) of the solvent-washed powder showed
427 a decrease in the proportion of oil compared to their parent oil powders, which was due
428 to the removal of free oil from the surface by the petroleum spirit. Moreover, the images
429 of the solvent-washed powders showed an increase in the proportion of oil globules on
430 the powder with the increase in total oil content in the powder (i.e. 40% fat). Such
431 correlation between the CLSM image of high-oil powder and surface fat content
432 indicates that CLSM imaging techniques can be utilized as a rapid quality control tool
433 during the production of plant-based high-oil powder.





434

435 **Figure 4.** CLSM images of oil powders and solvent-washed high-oil powders. a) 20% fat
436 content powder from 0.47 μm parent emulsion, b) 20% fat content powder from 0.75 μm
437 parent emulsion, c) 40% fat content powder from 0.47 μm parent emulsion, d) 40% fat



438 content powder from 0.75 μm parent emulsion, e) Solvent washed 20% fat content powder
439 from 0.47 μm parent emulsion, f) Solvent washed 20% fat content powder from 0.75 μm
440 parent emulsion, g) Solvent washed 40% fat content powder from 0.47 μm parent emulsion,
441 h) Solvent washed 40% fat content powder from 0.75 μm parent emulsion. Green dots
442 represent oil globules present on the powder surface.

443

444 3.7. Scanning electron microscopy (SEM) of corn oil powders

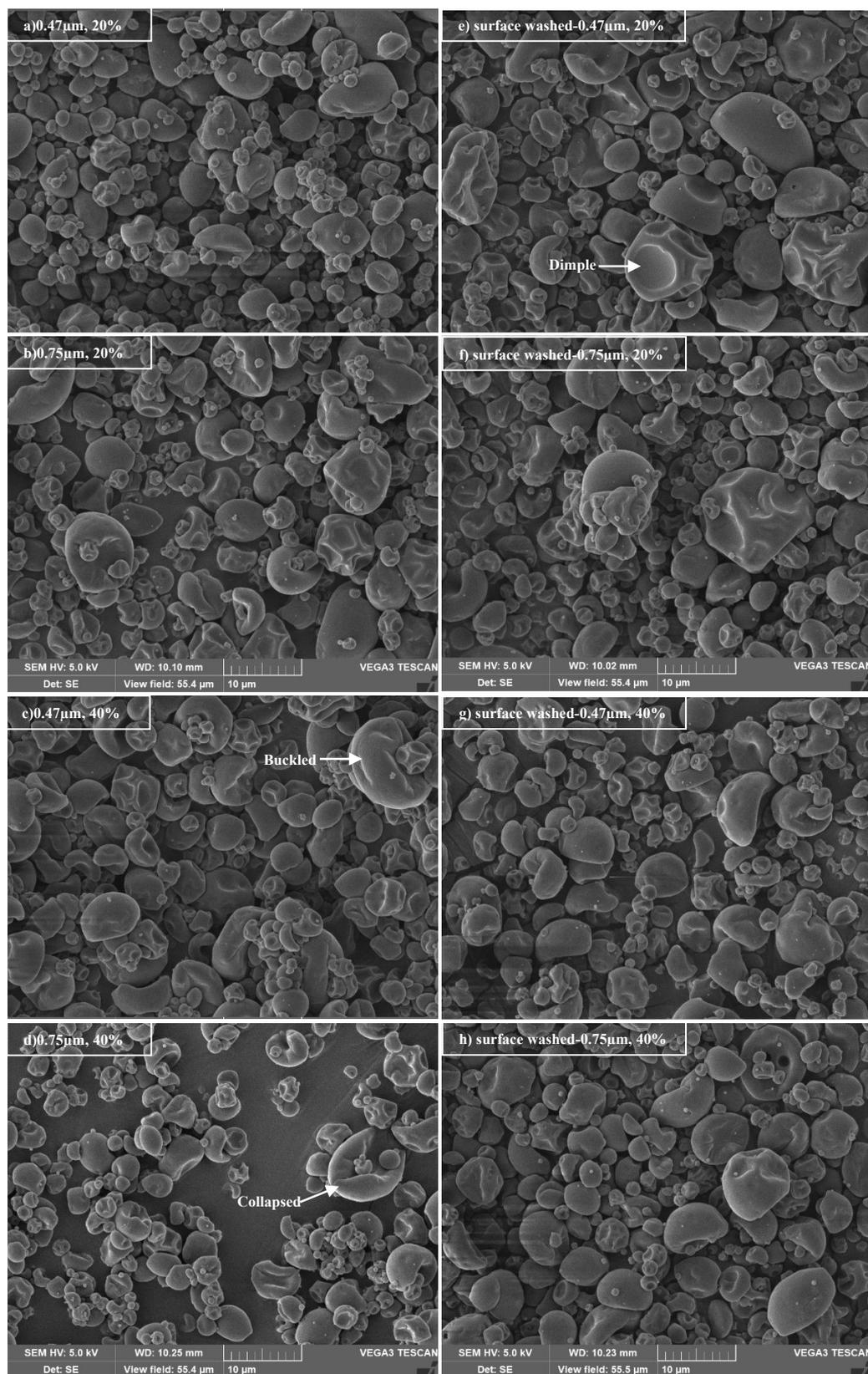
445 The surface structure of the oil-encapsulated powders was studied using scanning
446 electron microscopy (Fig. 5). The total field captured in each image is 50 μm . Overall,
447 it is evident from the images (Fig. 5, a, b, c, &d) of samples without surface washing
448 that there are lesser number of particle clusters. However, Cluster formation in oil/fat
449 encapsulated powder is primarily influenced by surface fat content in the powder [33].
450 In this present study, based on powder weight (in the section 3.3, free oil is expressed
451 based on total oil weight in powder), the free oil content of the powder samples, PS0.75-
452 40, PS0.47-40, PS0.75-20 and PS0.47-20 were 11.80 ± 0.14 , 9.32 ± 0.12 , 4.44 ± 0.10 and
453 3.92 ± 0.04 %, respectively. These free oil content values are relatively small and are
454 reflected in the prevalence of a large proportion of individual particles (Fig. 5, a, b, c,
455 &d) in bulk powder, which can further be correlated to the oil globule size distribution
456 of emulsions before spray drying and after reconstitution of powder (in section 3.5).
457 Therefore, it is very clear that the preparation of plant-based high-oil containing powder
458 with superior dispersibility and recosistitutability is possible with the method reported in
459 this study. There are very limited reports on production of such powder.
460 Further, the particles in high-oil powder having 20% oil content prepared from a parent
461 emulsion size 0.47 μm , had a comparatively rounder surface structure (Fig. 5a) than the
462 other high-oil powders (Fig. 5 b, c & d), which is reflected as better flow properties or



463 lower angle of repose. A rounder particle structure results in better flowability of the
464 powders[38]. The flowability of powder significantly affects the ease of handling during
465 industrial processes such as filling packs. In those images, it is evident that there is an
466 increase in the number of dimpled powder particles with an increase in the average oil
467 globule size of the parent emulsion. During spraying, the bigger oil globules tend to
468 migrate to the surface of the spray droplets. Besides, the oil globules on the surface
469 experience more physical stress during drying. As the bigger oil globules are sensitive
470 to physical stress, they either buckle or rupture during drying. Furthermore, the increase
471 in total oil content in the powder increased the number of buckled and dimpled particles.
472 An increase in oil phase decreases the wall material-to-oil ratio, resulting in a thinner
473 layer of wall material surrounding the oil globules. Such a condition also leads to
474 collapsing, buckling and dimple formation, especially in larger oil droplets. Similar
475 images were also obtained by Kim et al.[39] and Jones et al.[40] in spray-dried powders
476 from dairy and soybean oil emulsion, respectively.

477 In addition, solvent washing of the plant-based high-oil powders disintegrated powder
478 clusters to a greater extent in each formulation, an expected phenomenon. The
479 disintegration of powder particles is because of the removal of surface oil that was acting
480 as a connector in powder particle clusters. Removal of surface oil exposed buckled oil
481 globules as well as dimples present in oil globules. These are visible in the SEM images
482 of the solvent-washed powders in Fig. 5e, f, g, & h)





483

484 **Figure 5.** SEM images of fat powders and solvent-washed fat powders. a) 20% fat content485 powder from 0.47 μm parent emulsion, b) 20% fat content powder from 0.75 μm parent

486 emulsion, c) 40% fat content powder from 0.47 μm parent emulsion, d) 40% fat content
487 powder from 0.75 μm parent emulsion, e) Solvent washed 20% fat content powder from 0.47
488 μm parent emulsion, f) Solvent washed 20% fat content powder from 0.75 μm parent
489 emulsion, g) Solvent washed 40% fat content powder from 0.47 μm parent emulsion, h)
490 Solvent washed 40% fat content powder from 0.75 μm parent emulsion.

491

492 **4. Conclusion**

493 The present study demonstrated that a plant-based high fat powder can be prepared using
494 all plant-based materials formulations. In this study, the soluble fraction of mung bean
495 protein isolate (supernatant; 0.3% w/w protein) was used as the emulsifier. Corn oil was
496 used as the oil phase, and maltodextrin was used as the wall material. The fat powders
497 with an average fat globule size ($D[4,3]$) of 0.47 μm and 0.75 μm and with a total fat
498 content of 20% and 40% were prepared. Fat powder prepared from the corn oil emulsion
499 with the lowest mean fat globule size i.e. ($D[4,3]$) of 0.47 μm and 20% total fat, was
500 found to have better physicochemical properties compared to other formulations. This
501 fat powder had the lowest angle of repose, better reconstitutability, higher bulk density,
502 and lower free fat content. Furthermore, an increase in total fat content in the powder
503 and mean fat globule size in the initial emulsion led to an increase in free fat content and
504 angle of repose of the fat powders. These findings were further supported by the CLSM
505 and SEM images. The results from the present study are useful in the development of
506 plant-based food products as well as in our transition towards a sustainable future.

507 Although this study demonstrated that mung bean protein isolate/corn oil/maltodextrin could
508 be used to prepare plant-based high-fat powders, further research on the other plant proteins,
509 oils and wall materials can be explored.

510



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515 **Conflict of interest**

516 All authors declare no conflict of interest associated with this submission.

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All the relevant data is already present in the manuscript. However, other supplementary data can be made available upon a valid request.

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